



## Acute and reproductive effects of Align®, an insecticide containing azadirachtin, on the grape berry moth, *Lobesia botrana*

F. Javier Sáenz-De-Cabezón Irigaray<sup>a</sup>, Fernando Moreno-Grijalba<sup>b</sup>, Vicente Marco<sup>c</sup>, and Ignacio Pérez-Moreno<sup>d</sup>

Unidad de Protección de Cultivos Departamento de Agricultura y Alimentación, Universidad de La Rioja, C/ Madre de Dios, 9 51, 26006-Logroño (La Rioja), Spain

### Abstract

Azadirachtin, derived from the neem tree, *Azadirachta indica* A. Juss (Sapindales: Meliaceae), seems promising for use in integrated pest management programs to control a variety of pest species. A commercial formulation of azadirachtin, Align®, has been evaluated against different developmental stages of the European grape berry moth, *Lobesia botrana* Denis and Schifferrmüller (Lepidoptera: Tortricidae). When administered orally, Align reduced the fecundity and fertility of adults treated with 1, 5, and 10 mg litre<sup>-1</sup>. At the highest doses, fecundity and fertility were zero, but longevity was not affected. An LC<sub>50</sub> of 231.5 mg litre<sup>-1</sup> was obtained when Align was sprayed on eggs less than 1 day old. Hatching of all egg classes was significantly reduced, and this reduction was more pronounced for eggs less than 24 h old. LC<sub>50</sub> values of 2.1 mg litre<sup>-1</sup> for first instars and 18.7 mg litre<sup>-1</sup> for third instars were obtained when Align was present in the diet. Larvae reared on a diet containing different concentrations of Align did not molt into adults at the highest concentrations (0.3, 0.6, 1.2), and 50% molted at the lowest concentration (0.15). Phenotypic effects included inability to molt properly and deformities. The combination of acute toxicity and low, effective concentrations of Align observed in this study could lead to the inclusion of insecticides containing azadirachtin in integrated management programs against this pest.

**Keywords:** larval mortality, egg mortality, adult sterility, sublethal doses

**Correspondence:** <sup>a</sup>francisco-javier.saenz-de-cabezón@unirioja.es, <sup>b</sup>fernando.moreno@alum.unirioja.es, <sup>c</sup>vicente.marco@unirioja.es, <sup>d</sup>ignacio.perez@daa.unirioja.es

**Associate Editor:** James Ottea was editor of this paper

**Received:** 24 February 2008, **Accepted:** 10 October 2008

**Copyright :** This is an open access paper. We use the Creative Commons Attribution 3.0 license that permits unrestricted use, provided that the paper is properly attributed.

**ISSN:** 1536-2442 | Vol. 10, Number 33

Cite this paper as:

Irigaray FJ, Moreno-Grijalba F, Marco V, Pérez-Moreno I. 2010. Acute and reproductive effects of Align®, an insecticide containing azadirachtin, on the grape berry moth, *Lobesia botrana*. 11pp. *Journal of Insect Science* 10:33 available online: [insectscience.org/10.33](http://insectscience.org/10.33)

## Introduction

---

The grape berry moth *Lobesia botrana* Denis and Schiffermüller (Lepidoptera: Tortricidae), is a major pest of European and Mediterranean vineyards. The yield reductions caused by this insect are due to both damage done by the larvae and further attack by fungi. Most growers control this pest with traditional chemical pesticides, however, mating disruption and microbiological insecticides are used as alternatives in a few areas (Coscollá 1997). Considerable effort is being directed toward reduced use of traditional pesticides and increased use of integrated pest management techniques, emphasizing the joint use of natural enemies and selective pesticides, an alternative compatible with the protection of non-target organisms and the environment.

Azadirachtin, a tetranotriterpenoid, is the most active insecticidal compound found in the seeds and leaves of the neem tree, *Azadirachta indica* A. Juss (Sapindales: Meliaceae) (Koul et al. 1990; Schmutterer 1990). This compound exerts several biological effects on insects (e.g. antifeedant, insect growth regulator and repellent) and has low mammalian toxicity (Schmutterer 1990; Ascher 1993; Mordue and Blackwell 1993; Mordue et al. 1998; Mulla and Su 1999; Koul and Wahab 2004; Isman 2006). Pesticides containing azadirachtin are reported to provide broad-spectrum control of more than 200 species of insect pests (Ascher 1993) and to be less toxic to natural enemies of insect pests compared with synthetic, chemical pesticides (Hoelmer et al. 1990; Schmutterer 1997). Although the safety of azadirachtin for numerous beneficial insects has been recently questioned (Spollen and Isman 1996), applications of formulated

azadirachtin had no undesired effects on important predators such as coccinellids (Banken and Stark 1998), lacewings (Medina et al. 2004) and Phytoseiid mites (Castagnoli et al. 2005; Brito et al. 2006). Thus, pesticides derived from the neem tree seem to be promising for use in integrated pest management programs to control various pest species (Isman 2006).

Grape growers use large amounts of traditional chemical pesticides, and, therefore, the development of strategies that minimize pesticide use is needed. According to the European Directive 98/8/EC, “proper pesticide use includes application at an efficacious concentration and minimization of use of biocidal products where possible.” To ensure sufficient management of pests, registered pesticides are normally applied at rates above the upper asymptote of the concentration/response curve. Thus, in theory, it should be possible to achieve control of insect pests and, at the same time, reduce the effects on non-target insects by using dosage rates below these maxima, with the species-specific degree of selectivity strongly influenced by the applied concentration (Poehling 1989). Published studies of dose reduction from recommended field rates demonstrated adequate levels of control and improved control exerted by natural enemies without sacrificing yield or inflating cost (Smith et al. 1985; Poehling 1989; Acheampong and Stark 2004).

In the only published study of azadirachtin on *L. botrana*, Ioriarti et al. (1992) obtained 30% mortality on 3-day-old eggs and 52.6% mortality on larvae, using doses of 500 – 600 mg litre<sup>-1</sup>. Studies are needed to determine the most efficient usage against this pest. Herein the effects of azadirachtin on adult

fecundity, fertility and longevity are described. This study also reports the effects of azadirachtin on the development and mortality of *L. botrana* eggs and larvae. Finally, the possibility of incorporating azadirachtin into programs for management of the European grape berry moth are discussed.

## Materials and methods

### Insects

A stock culture of *L. botrana* was established from larvae collected in an organic vineyard in La Rioja, Spain during May 2000, and the culture was augmented with new individuals once each year. The insects were maintained in a growth chamber at  $24\pm 1^\circ\text{C}$ ,  $60\pm 10\%$  RH and 16:8 L:D photoperiod, following the method described by Del Tío (1996) and modified by Sáenz-de-Cabezón (2003). All bioassays were conducted under these uniform conditions.

### Chemicals

Formulated azadirachtin (ALIGN® EC, 32 g. azadirachtin litre<sup>-1</sup>) was obtained in Spain from Sipcam Inagra.

### Effect on adult fecundity, fertility and longevity

To assess the activity of azadirachtin on *L. botrana* adults, three pairs of recently emerged adults were introduced into a 33 ml plastic container placed upside-down on the base of a 90 mm diameter Petri dish. Azadirachtin was administered orally via a water trough that contained 1, 5, and 10 mg litre<sup>-1</sup> of azadirachtin mixed in a 10% honey solution. Water troughs were changed every 3 days to avoid fungal proliferation. Every day, the glasses with the laid eggs were replaced, and the eggs were then transferred

to plastic boxes (12 cm diameter, 5 cm high) until emergence. Fecundity and fertility were counted daily during the oviposition period. Mortality was recorded during the entire life period. Five replicates were used for both the treatment and the control (10% honey solution). Embryonic development was followed using a stereoscopic lupe (Olympus SZH-10).

### Ovicidal bioassays

To test topical ovicidal effects, ten pairs of *L. botrana* adults were introduced into an oviposition chamber consisting of a cylindrical plastic body (9 cm diameter, 13 cm high) covered inside with a transparent plastic film on which females could lay their eggs. The bottom was covered with a Petri dish, and the top was covered with filter paper. Moths were provided with water. Eggs laid during the next 24h were removed and then treated using a manually loaded Potter spray tower, with 5.5 ml in the tank at 20 kPa pressure producing a deposit of  $0.05\pm 0.01$  ml cm<sup>-2</sup>. The concentrations that were used were obtained by preliminary assays ranging from 1 to 768 mg litre<sup>-1</sup>. Eggs of different age classes (0-24, 24-48, 48-72 and 72-96 h old) were collected and treated in the same manner. Treatments were made on each of the age groups. The concentration used was the LC<sub>50</sub> obtained in the previous assay. Egg hatch was recorded 5 days after treatment. There were five replicates per concentration/treatment, in addition to the water treated control.

Ovicidal effects of Align were also examined using a contact bioassay. Surfaces were treated by dipping four plastic glasses in a 1:1 solution of LC<sub>50</sub> contact concentration, and then air-drying for 5 min. Five pairs of adults were introduced and allowed to lay eggs for 24 h. Larval

emergence from eggs was checked at the sixth day of treatment. Four replicates were used for the treatment and the control, which was dipped in distilled water.

### Larvicidal bioassays

To determine the effect of Align on the 1st, 3rd and 5th instars, ten larvae were introduced to a Petri dish (5 cm diameter) containing treated diet at concentrations ranging from 0.75 to 3.8 mg litre<sup>-1</sup> for 1<sup>st</sup> instars and 8.0 to 32.0 mg litre<sup>-1</sup> for 3rd instars. The concentrations used were established by preliminary assays. 5th instars were not treated because of delayed effects in development at  $\geq 30$  mg litre<sup>-1</sup>. Five replicates were used for each dose and control. The mortality (larvae that failed to molt) was measured when control larvae reached the next stadium (i.e., the 4th and 3rd day after treatment for 1st and 3rd instars, respectively). Sublethal effects of Align exposure were measured during the larval stage by providing 1st instar larvae *ad libitum* diet containing low concentrations of Align (i.e. 0.15, 0.3, 0.6, 1.2 mg litre<sup>-1</sup>). Mortality and adult emergence were checked daily. Five replicates, of 10 individuals each,

were used for each concentration and the control.

### Statistical analysis

Estimates of LC<sub>50</sub> and LC<sub>90</sub> for different stages and their 95% fiducial limits were obtained using the POLO program (Russell et al. 1977) based on Finney (1971). The criterion used to estimate the differences between LC ratios was non-overlap of their 95% confidence intervals (Robertson and Presley 1992). The significance of results (fecundity, fertility, egg mortality, and larval mortality) were tested by ANOVA, and means were separated by an LSD multiple range test ( $p < 0.05$ ). Percentage values were arcsin transformed, and, in all cases, untransformed means are presented. Abbott's formula was used to correct mortalities (Abbott 1925). Percentage of sterility was calculated using the formula of Topozada et al. (1966):

$$\% \text{ Sterility} = [1 - (F_t \times F_e / F_c \times F_{e_c})] \times 100$$

where  $F_t$  = fecundity of treated females;  $F_e$  = fertility of treated females;  $F_c$  = control fecundity; and  $F_{e_c}$  = control fertility

**Table 1.** Fecundity and fertility parameters of adult *L. botrana* pairs treated *ad libitum* with different concentrations of azadirachtin 1, 5, and 10 mg litre<sup>-1</sup>

Doses (mg litre)	n	Fecundity $\pm$ SD	Percentage of fertility	Corrected fertility (Abbott)	Sterility percentage
0	5	163.7 $\pm$ 18.45 a	89.9 $\pm$ 4.44a	-	-
1	5	117.9 $\pm$ 21.18b	66.0 $\pm$ 6.26b	26.6 $\pm$ 6.96	47.1
5	5	0.0 $\pm$ 0.0c	0 $\pm$ 0.0c	100	100
10	5	0.0 $\pm$ 0.0c	0 $\pm$ 0.0c	100	100

Means followed by different letters are significantly different at  $\alpha = 0.05$ . Every mean is a result of three replicates

## Results

### Effects on adults

*L. botrana* adults fed Align in a honey solution showed decreased fecundity and fertility at each concentration tested (Table 1). Total eggs laid per female were significantly ( $p < 0.05$ ) less than the control, and fertility was significantly ( $p < 0.05$ ) decreased in *L. botrana* adults. In addition, eggs laid by Align-treated females showed the same toxic symptoms on the embryo as spray-treated eggs (see below). Longevity was not affected by the treatment of any concentration of Align.

### Effects on eggs

The parameters obtained for the probit-log concentration regression line on eggs treated with azadirachtin and less than 24 h are given in Table 2. Contact treatment was significantly ( $p < 0.05$ ) more effective than surface treatment (with  $41.4 \pm 4.3$  and  $30.0 \pm 7.1$  corrected mortalities, respectively). Treatment with Align significantly ( $p < 0.05$ ) reduced hatching for all egg classes but the last (72-96 h old). The reduction was greater for eggs less than 24 h old than for other egg classes (Table 3). High concentrations of Align ( $768 \text{ mg litre}^{-1}$ ) interrupted the normal development of the embryo. Treated eggs reached the final black-head stage but had mandibular and cephalic deformities.

### Effects on larvae

Table 2 shows probit-log concentration regression line parameters obtained for the different stages tested. Repellent effects of Align were evident during the course of preliminary assays, when death due to starvation was evident in feeding experiments with concentrations higher than  $12 \text{ mg litre}^{-1}$ . Younger larvae were more susceptible than older larvae. Effects on larval development were observed and included the inability to shed the old cuticle and cephalic capsule. In addition, larvae that successfully molted exhibited mouthpart deformities, integument injuries and malformations. Treatment of 5th instars with concentrations higher than  $32 \text{ mg litre}^{-1}$  of Align caused larvae to prematurely enter a wandering state and molt the pupal stage. Hardening the cuticle sclerites and other abnormalities, such as larval-pupal intermediates, were also observed. When treated with reduced concentrations of Align, there was a high percentage of larval mortality in the 5th stadium for all concentrations tested except  $0.15 \text{ mg litre}^{-1}$ . Mortality on the next stadia reached  $>85\%$  in all concentrations but  $0.15 \text{ mg litre}^{-1}$ . Almost 100% of the individuals died without reaching adulthood at concentrations above  $0.15 \text{ mg litre}^{-1}$ . The lowest concentration reduced adult elision up to 50%. Mortality occurred mostly during the last larval stage (Figure 1).

**Table 2.** Probit-log concentration-response regression line parameters, of different stages of *L. botrana* treated with azadirachtin.

Stadium	Slope $\pm$ SE	Intercept $\pm$ SE	$\chi^2$	g	LC <sub>50</sub>	LC <sub>90</sub>
					(C.L. 95%)	(C.L. 95%)
Egg	0.491 $\pm$ 0.103	3.84 $\pm$ 0.228	0.53	0.169	231.5 (95.9; 782.2)	94,799 (11271; 1.2E+7)
L <sub>1</sub>	2.207 $\pm$ 0.353	4.278 $\pm$ 0.121	0.08	0.098	2.125 (1.788; 2.267)	8.089 (5.460; 16.689)
L <sub>3</sub>	2.475 $\pm$ 0.509	1.851 $\pm$ 0.611	2.7	0.163	18.71 (16.08; 23.49)	61.65 (40.56; 160.82)

Doses are given in mg litre<sup>-1</sup>

## Discussion

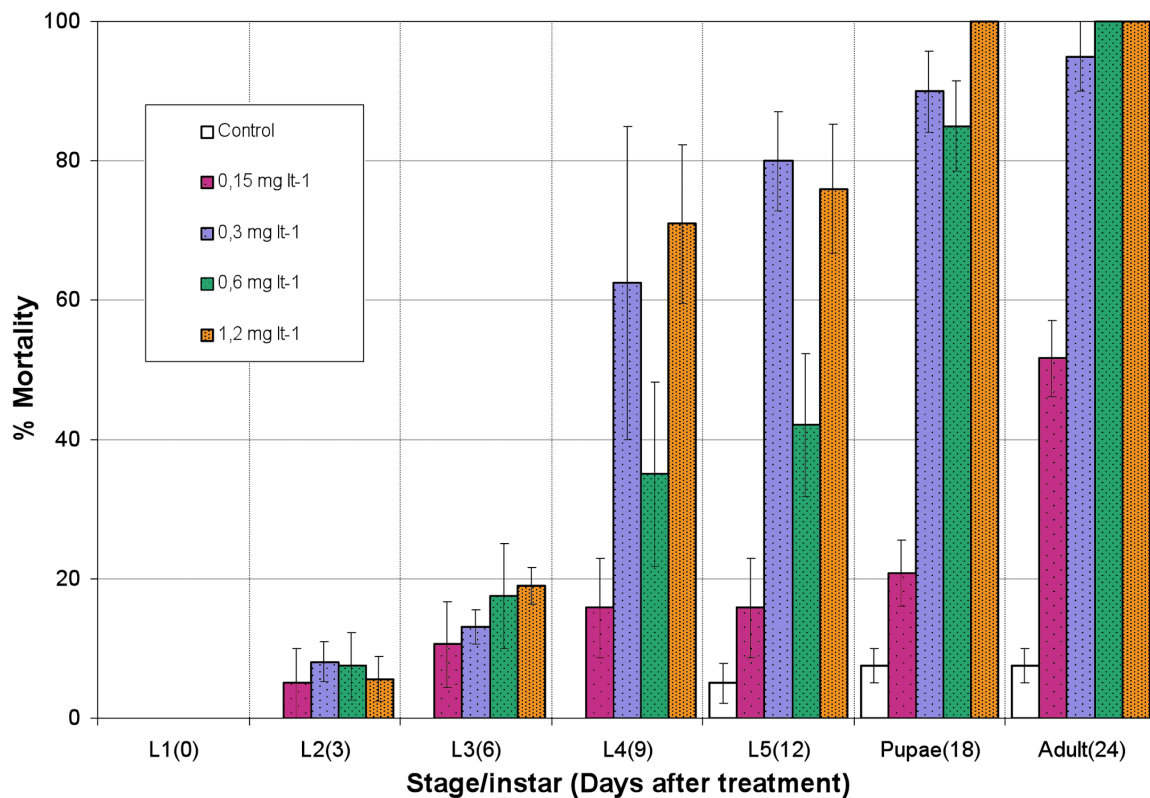
Align®, a commercial formulation of azadirachtin, has larvicidal and ovicidal activity and significantly affected the reproduction of *L. botrana*. At low concentrations total adult eclosion was reduced to 0%.

In a previous study with this insect, Iorriati et al. (1992) measured mortalities of 30% for 3-day-old eggs and 52.6% for larvae using concentrations between 500 and 600 mg litre<sup>-1</sup>. In the present study the effects of Align on eggs, larvae and adult *L. botrana* have been assessed, emphasizing the time and mode of treatment.

**Table 3.** Hatching percentage and corrected egg mortality of four age classes of *L. botrana* treated with 231.5 mg litre of azadirachtin.

Egg (hours)	n	Corrected mortality (Abbott)	
		Control	Treated
		0 – 24	-
24 – 48	5	-	16.0 ± 4.0b
48 – 72	5	-	14.0 ± 2.5b
72 – 96	5	-	-

Means followed by different letters are significantly different at  $\alpha = 0.05$ .



**Figure 1.** Stage-instar mortality when 1<sup>st</sup> instar larvae of *Lobesia botrana* are treated *ad libitum* during the larval stage with low concentrations of azadirachtin (mg litre). High quality figures are available online.

### **Effects on adult fertility, fecundity, and longevity**

Align strongly affected the reproduction of *L. botrana* by producing a high percentage of sterility at the concentrations tested. Fecundity was severely reduced, and no eggs were laid at the highest concentrations tested. Lower concentrations significantly reduced the fertility of *L. botrana*. Eggs laid by treated females showed embryocidal effects, which suggest pesticide transference through the gravid female. It has been demonstrated that adult exposure to azadirachtin by topical, surface or ingestion treatments, reduces fecundity and/or fertility among different insect orders, including Lepidoptera (Schmutterer 1990; Ascher 1993; Mordue and Blackwell 1993; Mordue 2004; Seljansen and Meadow 2006). It is known that azadirachtin causes profound effects on reproductive processes of both male and female insects. Studies showed that insects treated with azadirachtin have degenerate ovaries and a high degree of yolk resorption (Rembold and Seiber 1981; Koul 1984; Dorn et al. 1987; Schlüter 1987; Medina et al. 2004). Azadirachtin also interferes with the synthesis of vitellogenin by the fat body and its uptake by the eggs, resulting in reduced fecundity and sterility. Such effects could be due to the disruption of juvenile hormone levels and ovarian ecdysteroid production (Rembold and Seiber 1981; Feder et al. 1988; Tanzubil and McCaffery 1990).

### **Ovicidal effects**

Align was not a potent ovicide of *L. botrana* eggs ( $LC_{50}$  of 231.5 mg litre<sup>-1</sup> for eggs < 24 h). Ovicidal activity was also affected by age and treatment. Younger eggs were more susceptible than older ones, and surface treatment was less effective than spray treatment. These findings are in agreement

with those from previous studies showing that azadirachtin has low ovicidal activity on *L. botrana* (Iorriati et al. 1992) and on insects of different orders (Schmutterer 1990; Ascher 1993). The results from the present study differ from previous studies (Ioriatti et al. 1992) that obtained a greater susceptibility on 3-day-old eggs.

### **Larvicidal effects**

*L. botrana* larvae were highly susceptible to diet containing Align, with younger stages being more susceptible than older ones. These results agree with those from previous studies (Mordue and Blackwell 1993) of other lepidopteran insects showing larvicidal activity inversely proportional to larval age. In addition to increased mortality, Align acted as an insect growth regulator and an antifeedant: 1st instars did not feed when concentrations were higher than 12 mg litre<sup>-1</sup> (data not shown) (Schmutterer 1990; Ascher 1993; Mordue and Blackwell 1993). Also, 5th instars displayed premature wandering and cessation of feeding, possibly due to Align's effects on release of prothoracicotropic hormone. As described for other species, an increase in the levels of prothoracicotropic hormone is necessary to begin the transformation into pupa. Without this increase, the larvae remain in a state of wandering and die (Schmutterer 1990; Mordue 2004). In a study on *Tripanosoma cruzi*, the complete elimination of azadirachtin did not occur until 50 days after the treatment. Thus, this compound can interrupt the insect development for a long time (González and García 1992). Low concentrations of Align resulted in elevated mortality during the last larval stage, probably as a result of compound accumulation reaching concentrations that have activity in older instars.

### Integrated Pest Management

The main impediment for the implementation of integrated pest management is to provide growers with a convincing reason to adopt its tactics, such as economic incentives (Reitz et al. 1999). Integrated pest management often increases net profits for growers who adopt it; however, many growers still hold the perception that it does not offer short-term economic advantages compared with conventional control. However, in a previous study (Sáenz-de-Cabezón 2003), Align was as effective as the conventional organophosphate fenitrothion, even when applied at a concentration (50 cc/hl) lower than that recommended on the label (125 cc/hl; data not shown). Thus, proper pesticide-use tactics reduce the cost of pesticide use and further the implementation of integrated pest management programs.

More laboratory, small plot and field testing is required before the insecticidal potential of azadirachtin against *L. botrana* will be fully understood. The activity observed in this study suggests that the combination of the acutely toxic and reduced concentrations could lead to the incorporation of this compound in integrated pest management programs against the grape berry moth. Sprays for the second and third generations should be applied at the beginning of flight, to cover the oviposition period and egg hatching. Resistance studies are needed in order to know how azadirachtin resistance works in *L. botrana* populations using different levels of selection pressure.

### Acknowledgements

The authors thank the Comisión Interministerial de Ciencia y Tecnología (CICYT) and Fondo Europeo de Desarrollo

Regional (FEDER) for funding this research. We also thank Christina Pease for reviewing the English version of the manuscript.

### References

Abbott WS. 1925. A method of computing the effectiveness of an insecticide. *Journal of Economic Entomology* 18: 265-267.

Acheampong S, Stark JD. 2004. Can reduced rates of pymetrozine and natural enemies control the cabbage aphid, *Brevicoryne brassicae* (Homoptera: Aphididae), on broccoli? *International Journal of Pest Management* 50: 275-279.

Ascher KRS. 1993. Nonconventional insecticidal effects of pesticides available from the neem tree, *Azadirachta indica*. *Archives of Insect Biochemistry and Physiology* 22: 433- 449.

Banken JAO, Stark JD. 1998. Multiple routes of pesticide exposure and the risk of pesticides to biological controls: A study of neem and seven-spotted lady beetle (Coleoptera: Coccinellidae). *Journal of Economic Entomology* 91: 1-6.

Brito HM, Gondim-Junior MGC, Oliveira JV de, Camara CAG da. 2006. Toxicity of neem (*Azadirachta indica* A. Juss) formulations for twospotted spider mite and *Euseius alatus* De Leon and *Phytoseiulus macropilis* (Banks) (Acari: Phytoseiidae). *Neotropical Entomology* 35: 500-505.

Castagnoli M, Nannelli R, Simoni S. 2005. Effects of repeated applications of an azadirachtin-based product on the spider mite *Tetranychus urticae* and its phytoseiid predator *Neoseiulus californicus*. *Bull OILB/SROP* 28: 51-54.

Coscollá R. 1997. *La polilla del racimo de la vid (Lobesia botrana Den. y Schiff.)*. Ed. Generalitat Valenciana-Conselleria de Agricultura, Pesca y Alimentación. Valencia.

Del Tío R. 1996. *Estudios Biológicos y modelos de predicción para el control integrado de Lobesia botrana Denis Schiffermüller (Lepidoptera Tortricidae) en el marco de Jerez*. Ph. D. dissertation. University of Sevilla. Spain.

Dorn A, Rademacher JM, Shen E. 1987. Effects of azadirachtin on reproductive organs and fertility in the large milkweed bug, *Oncopeltus fasciatus*, in Natural Products from the Neem tree and Other Tropical Plants, pp. 273-288. In: H Schmutterer and KRS Ascher, editors. *Proceedings 3<sup>rd</sup> International neem Conference*, GTZ, Eschborn, Germany.

Feder D, Valle D, Rembold H, García SE. 1988. Azadirachtin-induced sterilization in mature females of *Rhodnius prolixus*. *Zeitschrift für Naturforschung* 43C: 908-903.

Finney DJ. 1971. *Probit Analysis*. Cambridge University Press.

González MS, García ES. 1992. Effect of azadirachtin on the development of *Tripanosoma cruzi* in different species of triatomine insect vectors: long-term and comparative studies. *Journal of Invertebrate Pathology* 60: 201-205.

Hoelmer KA, Osborne LS, Yokomi RK. 1990. Effects of neem extracts on beneficial insects in greenhouse culture. In: J. C. Locke R. H. Lowson, editors. *Neem's potential in pest management programs*, pp. 100-105. USDA Agricultural Research Service, 86.

Ioriatti C, Dalri M, Delaiti M, Delaiti L. 1992. Studio dell'attività e della persistenza d'azione di alcuni regolatori di crescita degli insetti su *Lobesia botrana* (Den. et Schiff.). *ATTI Giornate Fitopatologiche* 1: 147-154.

Isman MB. 2006. Botanical insecticides, deterrents, and repellents in modern agriculture and an increasingly regulated world. *Annual Review of Entomology* 51: 45-66.

Koul O. 1984. Azadirachtin I. Interaction with the development of red cotton bugs. *Entomologia Experimentalis et Applicata* 36: 85-88.

Koul O, Isman MB, Retkar CM. 1990. Properties and uses of neem, *Azadirachta indica*. *Canadian Journal of Botany* 68: 1-11.

Koul O and Wahab S. 2004. *Neem: today and in the new millennium*. Kluwer Academic Publishers.

Medina P, Budía F, Del Estal P, Viñuela E. 2004. Influence of azadirachtin, a botanical insecticide, on *Chrysoperla carnea* (Stephens) reproduction: toxicity and ultrastructural approach. *Journal of Economic Entomology* 97: 43-50.

Mordue AJ. 2004. Present concepts of the mode of action of azadirachtin from Neem. In: Koul O, Wahab S, editors. *Neem: Today and in the New Millennium*. pp. 229-242. Kluwer Academic Publishers.

Mordue AJ, Blackwell A. 1993. Azadirachtin: an update. *Journal of Insect Physiology* 39: 903-924.

Mordue AJ, Simmonds MSJ, Ley SV, Blaney WM, Mordue W, Nasiruddin M and Nisbet AJ. 1998. Actions of azadirachtin, a

plant allelochemical, against insects. *Pesticide Science* 54: 277-284.

Mulla MS, Su T. 1999. Activity and biological effects of neem products against arthropods of medical and veterinary importance. *Journal of the American Mosquito Control Association* 14: 204-209.

Poehling HM. 1989. Selective application strategies for insecticides in agricultural crops. In: Jepson PC, editor. *Pesticides and Non-target Invertebrates*, pp. 151-175. Intercept.

Rembold H, Seiber KP. 1981. Inhibition of oogenesis and ovarian ecdysteroid synthesis by azadirachtin in *Locusta migratoria migratorioides* (R&F). *Zeitschrift für Naturforschung* 36C: 466-469.

Robertson JL and Preisler HK. 1992. *Pesticide Bioassays with Arthropods*. CRC Press.

Russell RM, Robertson JL, Savin NE. 1997. *POLO: a new computer program for Probit analysis*. Bulletin of the Entomological Society of America 23: 209-213.

Sáenz de Cabezón Irigaray FJ. 2003. *Aportaciones al manejo integrado de Lobesia botrana (Den. y Schiff.) (Lepidoptera: Tortricidae): evaluación de la eficacia de lufenurón, metoxifenocida y azadiractina y modelización del desarrollo*. Ph. D. Dissertation. Universidad de La Rioja. Spain.

Schlüter U. 1987. Effects of azadirachtin on developing tissues of various insect larvae, in Natural Products from the Neem tree and Other Tropical Plants. In: H. Schmutterer and K.R.S. Ascher., editors. *Proceedings 3<sup>rd</sup> International neem Conference*. pp. 331-348. GTZ, Eschborn, Germany.

Schmutterer H. 1985. Which insect pests can be controlled by application of neem seed kernel extracts under field conditions? *Zeitschrift für Angewandte Entomologie* 100: 468-475.

Schmutterer H. 1990. Properties and potential of natural pesticides from the neem tree, *Azadirachta indica*. *Annual Review of Entomology* 35: 271-297.

Schmutterer H. 1997. Side effects of neem (*Azadirachta indica*) products on insect pathogens and natural enemies of spider mites and insects. *Journal of Applied Entomology* 121: 121-128.

Seljansen R, Meadow R. 2006. Effects of neem on oviposition and egg and larval development of *Mamestra brassicae* L: dose response, residual activity, repellent effect and systemic activity in cabbage plants. *Crop Protection* 25: 338-345.

Smith RG, Archer TL, Bynum ED. 1985. Low-rate applications of insecticides for greenbug (Homoptera: Aphididae) control and effect on beneficial insects on sorghum. *Journal of Economic Entomology* 78: 988-991.

Spollen KM and Isman MB. 1996. Acute and sublethal effects of a neem insecticide on the commercial biocontrol agents *Phytoseiulus persimilis* and *Amblyseius cucumeris* (Acari: Phytoseiidae), and *Aphilodetes aphidimyza* (Rondani) (Diptera: Cecidomyiidae). *Journal of Economic Entomology* 89: 1379-1386.

Tanzubil PB, McCaffery AR. 1990. Effects of azadirachtin in the African armyworm (*Spodoptera exempta*). *Entomologia Experimentalis et Applicata* 57: 115-121.

Topozada A Abdallah S, Eldefrawi ME. 1966. Chemosterilization of larvae and adults of the egyptian cotton leafworm *Prodenia litura*, by apholate, metepa, and tepa. *Journal of Economic Entomology* 59: 1125-1128.